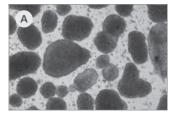
REPRODUCIBLY PRODUCE UNIFORM EMBRYOID BODIES AggreWell™ Plates

Many pluripotent stem cell (PSC) differentiation protocols begin with the formation of 3-dimensional aggregates of cells called embryoid bodies (EBs). EB size directly affects subsequent differentiation trajectories¹⁻⁷, but conventional EB formation methods^{8,9} result in EBs that are heterogeneous in size and shape, leading to inefficient and uncontrolled differentiation¹.

AggreWell™ plates provide an easy and standardized approach to the production of EBs. Each well contains microwells of defined size, making it easy to produce large numbers of highly uniform EBs and ensure reproducibility of differentiation experiments.



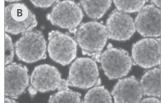
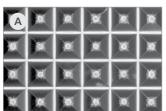


Figure 1. Generate Uniform Embryoid Bodies Using AggreWell™

(A) Human EBs formed using conventional methods are heterogeneous in size and shape resulting in inefficient differentiation. (B) Human EBs formed using AggreWell™ plates are uniform in size and consistently spherical in shape. Shown are EBs generated with 2,000 cells using AggreWell™400.

EBs and other cell aggregates¹⁰⁻¹² generated using AggreWellTM plates are highly uniform in size and shape, and consistent within and between experiments. EB size can be easily modified by adjusting the cell seeding density. EBs formed using AggreWellTM can be efficiently differentiated into a variety of cell types.



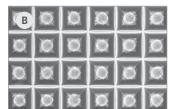
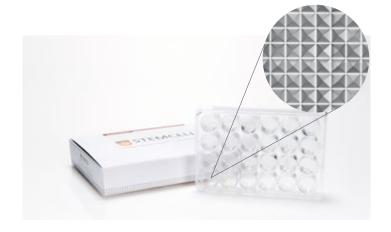


Figure 2. hPSCs Form Embryoid Bodies in AggreWell™ Plates

Starting from a single-cell suspension, hPSCs form uniform EBs after 24 hours in AggreWell™. The size of the EB can be easily modified by adjusting the seeding density. Shown are EBs in AggreWell™400 plates (A) 100 cells per microwell and (B) 1000 cells per microwell.



Why Use AggreWell™?

EASY TO USE. Simple EB generation.

REPRODUCIBLE. Large numbers of uniform EBs.

CONTROL OF EB SIZE. 50 to 20,000 cells per EB.

CONSISTENCY. Reduces variability in differentiation protocols.

HIGH YIELD. Up to 5,900 EBs per well.

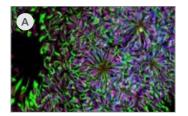




Figure 3. EBs Generated Using AggreWell™ Can Differentiate into Multiple Cell Types

(A) Neural progenitor cells (NPCs) were derived from EBs formed using AggreWell™800 plates and differentiated using STEMdiff™ Neural Induction Medium (Catalog #05835). NPCs express SOX1 (red), Nestin (green) and PAX6 (not shown), but not SOX10 (not shown). Nuclei are counterstained with DAPI. (B)* Hematopoietic progenitor cells were derived from EBs formed using AggreWell™400 plates and differentiated in serum-containing medium in suspension culture. Hematopoietic colony-forming units (CFUs) were detected using MethoCult™ H4435 Enriched (Catalog #04435).

*Data reprinted from Ungrin et al., 2008. See reference for full culture details.

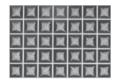


Formation of EBs in AggreWell™













Uniform EBs are ready for downstream use

AggreWell™ Products

suspension

Product	Size of Microwell	Plate Format	Size of EB	Number of EBs	Quantity	Catalog #
AggreWell™400	400 μm	24-well plate	50 - 3,000 cells	~ 1,200 per well	1/pack	34411
					5/pack	34415
		6 well plate		~ 5,900 per well	1/pack	34421
		6-well plate			5/pack	34425
AggreWell™800	800 µm	24-well plate	3,000 - 20,000 cells	~ 300 per well	1/pack	34811
		24-weii piate			5/pack	34815
		6-well plate		~ 1,500 per well	1/pack	34821
					5/pack	34825
AggreWell™HT	900 μm	96-well plate	50 - 20,000 cells	~32 per well	1/pack	200-0563
					5/pack	200-0570
AggreWell™ EB Formation Medium	Defined, serum-free medium for generation and culture of EBs using AggreWell™ plates				100 mL	05893
AggreWell™ Rinsing Solution†	Rinsing solution for AggreWell™ plates to reduce surface tension and ensure optimal EB formation				100 mL	07010
37 μm Reversible Strainers, Small	37 μm nylon mesh filter, fits standard 14 mL round bottom tubes & 15 mL conical tubes				20/box	27215
37 μm Reversible Strainers, Large	37 µm nylon mesh filter, fits standard 50 mL conical tubes				12/box	27250

Centrifuge gently to settle

the cells into the microwells

For a complete list of related products for hPSC culture and differentiation, including specialized cell culture and storage media, matrices, antibodies, cytokines, and small molecules, visit www.stemcell.com/hPSCworkflow or contact us at techsupport@stemcell.com.

References

- Bauwens CL et al. (2008) Control of human embryonic stem cell colony and aggregate size heterogeneity influences differentiation trajectories. Stem Cells 26: 2300–10.
- Ungrin MD et al. (2008) Reproducible, ultra-high-throughput formation of multicellular organization from single cell suspension-derived human embryonic stem cell aggregates. PLoS One 3(2): e1565.
- 3. Bratt-Leal AM et al. (2009) Engineering the embryoid body microenvironment to direct embryonic stem cell differentiation. Biotechnology Progress 25: 43–51.
- Hwang YS et al. (2009) Microwell-mediated control of embryoid body size regulates embryonic stem cell fate via differential expression of WNT5a and WNT11. Proc Natl Acad Sci USA 106: 16978–83.
- Messana JM et al. (2008) Size of the embryoid body influences chondrogenesis of mouse embryonic stem cells. J Tissue Eng Regen Med 2: 499-506.
- Mohr JC et al. (2010) The microwell control of embryoid body size in order to regulate cardiac differentiation of human embryonic stem cells. Biomaterials 31:1885–93.

- 7. Ng ES et al. (2005) Forced aggregation of defined numbers of human embryonic stem cells into embryoid bodies fosters robust, reproducible hematopoietic differentiation. Blood 106: 1601–03.
- Itskovitz-Eldor J et al. (2000) Differentiation of human embryonic stem cells into embryoid bodies compromising the three embryonic germ layers. Mol Med 6: 88–95.
- Kurosawa H. (2007) Methods for inducing embryoid body formation: in vitro differentiation system of embryonic stem cells. J Biosci Bioeng 103: 389–98.
- Razian G et al. (2013) Production of large numbers of size-controlled tumor spheroids using microwell plates. J Vis Exp. 81: e50665.
- Wallace L et al. (2013) Using 3D culture to investigate the role of mechanical signaling in keratinocyte stem cells. Methods Mol Biol 989: 153–64.
- Markway BD et al. (2010) Enhanced chondrogenic differentiation of human bone marrow-derived. Mesenchymal stem cells in low oxygen environment micropellet cultures. Cell Transplantation. 19: 29–42.

Copyright © 2025 by STEMCELL Technologies Inc. All rights reserved including graphics and images. STEMCELL Technologies & Design, STEMCELL Shield Design, Scientists Helping Scientists, and AggreWellTM are trademarks of STEMCELL Technologies Inc. All other trademarks are the property of their respective holders.

UNLESS OTHERWISE STATED, PRODUCTS ARE FOR RESEARCH USE ONLY AND NOT INTENDED FOR HUMAN OR ANIMAL DIAGNOSTIC OR THERAPEUTIC USES. FOR PRODUCT-SPECIFIC COMPLIANCE AND INTENDED USE INFORMATION, REFER TO THE PRODUCT INFORMATION SHEET. GENERAL INFORMATION ON QUALITY AT STEMCELL MAY BE FOUND AT WWW.STEMCELL.COM/COMPLIANCE.



[†] Required for optimal performance.