



## EasySep™ Human CD25 Positive Selection Kit

Positive Selection

Catalog #18231

For processing  $1 \times 10^9$  cells



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Document #28985 | Version 2\_4\_0

## Description

Isolate highly purified CD25+ cells by immunomagnetic positive selection from fresh or previously frozen human peripheral blood mononuclear cells (PBMCs) or samples that have been pre-enriched for CD4+ T cells or regulatory T cells.

- Fast and easy-to-use
- No columns required

This kit targets CD25+ cells for positive selection with an antibody recognizing the CD25 surface marker. Desired cells are labeled with antibodies and magnetic particles, and separated without columns using an EasySep™ magnet. Unwanted cells are simply poured off, while desired cells remain in the tube. Isolated cells are immediately available for downstream applications such as flow cytometry, cell culture, or DNA/RNA extraction.

- If isolating CD25+ cells as part of STEMCELL's Complete Kits for Human Regulatory T Cells (Catalog #15861, 15862, or 15864), refer to the applicable Product Information Sheet.

## Component Descriptions

| COMPONENT NAME                                     | COMPONENT # | QUANTITY | STORAGE                          | SHELF LIFE                               | FORMAT  |
|--|-------------|----------|----------------------------------|--|---|
| EasySep™ Human CD25 Positive Selection Cocktail    | 18231C.2    | 1 x 1 mL | Store at 2 - 8°C. Do not freeze. | Stable until expiry date (EXP) on label. | A combination of monoclonal antibodies in PBS and 0.1% BSA. |
| EasySep™ Magnetic Nanoparticles Positive Selection | 18150       | 1 x 1 mL | Store at 2 - 8°C. Do not freeze. | Stable until expiry date (EXP) on label. | A suspension of magnetic particles in water.                |

BSA - bovine serum albumin; PBS - phosphate-buffered saline

Components may be shipped at room temperature (15 - 25°C) but should be stored as indicated above.

## Sample Preparation

For available fresh and frozen samples, see [www.stemcell.com/primarycells](http://www.stemcell.com/primarycells).

### PERIPHERAL BLOOD

Prepare a PBMC suspension from whole blood by centrifugation over a density gradient medium (e.g. Lymphoprep™, Catalog #07801). For more rapid PBMC preparation, use the SepMate™ RUO (Catalog #86450/86415) or SepMate™ IVD\* (Catalog #85450/85415) cell isolation tube.

If using previously frozen PBMCs, incubate the cells with DNase I Solution (Catalog #07900) at a concentration of 100 µg/mL at room temperature (15 - 25°C) for at least 15 minutes prior to labeling and separation. Filter aggregated suspensions through a 40 µm Cell Strainer (Catalog #27305) for optimal results.

After preparation, resuspend cells at  $1 \times 10^8$  cells/mL in recommended medium.

\* SepMate™ IVD is only available in select regions where it is registered as an In Vitro Diagnostic (IVD) device for the isolation of mononuclear cells (MNCs) from whole blood or bone marrow by density gradient centrifugation.



## Recommended Medium

EasySep™ Buffer (Catalog #20144), RoboSep™ Buffer (Catalog #20104), or PBS containing 2% fetal bovine serum (FBS) and 1 mM EDTA. Medium should be free of Ca++ and Mg++.

## Directions for Use – Manual EasySep™ Protocols

See page 1 for Sample Preparation and Recommended Medium. Refer to Tables 1 and 2 for detailed instructions regarding the EasySep™ protocol.

**Table 1. EasySep™ Human CD25 Positive Selection Kit Protocol**

|      |  | EASYSEP™ MAGNETS   |  |
|------|--|--|--|
| STEP | INSTRUCTIONS   |  <b>EasySep™ (Catalog #18000)</b> | <b>“The Big Easy” (Catalog #18001)</b>              |
| 1    | Prepare sample at the indicated cell concentration within the volume range.  | 1 x 10 <sup>8</sup> cells/mL<br>0.1 - 2.5 mL   | 1 x 10 <sup>8</sup> cells/mL<br>0.25 - 8.5 mL  |
|      | Add sample to required tube.   | 5 mL (12 x 75 mm) polystyrene round-bottom tube<br>(e.g. Corning Catalog #352058)                                  | 14 mL (17 x 100 mm) polystyrene round-bottom tube<br>(e.g. Corning Catalog #352057)  |
| 2    | Add Selection Cocktail to sample.  | 50 µL/mL of sample   | 50 µL/mL of sample   |
|      | Mix and incubate.  | RT for 15 minutes  | RT for 15 minutes  |
| 3    | Mix Magnetic Particles.  | Pipette up and down more than 5 times  | Pipette up and down more than 5 times  |
| 4    | Add Magnetic Particles to sample.  | 50 µL/mL of sample   | 50 µL/mL of sample   |
|      | Mix and incubate.  | RT for 10 minutes  | RT for 10 minutes  |
| 5    | Add recommended medium to top up the sample to the indicated volume. Mix by gently pipetting up and down 2 - 3 times.  | Top up to 2.5 mL   | <ul style="list-style-type: none"> <li>• Top up to 5 mL for samples ≤ 4 mL</li> <li>• Top up to 10 mL for samples &gt; 4 mL</li> </ul> |
|      | Place the tube (without lid) into the magnet and incubate.   | RT for 5 minutes   | RT for 5 minutes   |
| 6    | Pick up the magnet, and in one continuous motion invert the magnet and tube,* pouring off the supernatant. Remove the tube from the magnet; this tube contains the isolated cells. | Discard supernatant<br>Or save for CD25 depletion protocol, see Table 2  | Discard supernatant<br>Or save for CD25 depletion protocol, see Table 2  |
| 7    | Repeat steps as indicated.   | Steps 5 and 6, three more times<br>(total of 4 x 5-minute separations)   | Steps 5 and 6, three more times<br>(total of 4 x 5-minute separations)   |
| 8    | Resuspend cells in desired medium. Be sure to collect cells from the sides of the tube.  | Isolated cells are ready for use   | Isolated cells are ready for use   |

RT - room temperature (15 - 25°C)



\* Leave the magnet and tube inverted for 2 - 3 seconds, then return upright. Do not shake or blot off any drops that may remain hanging from the mouth of the tube.

## Directions for Use – Manual EasySep™ Depletion Protocol

Following step 6 of a Manual EasySep™ Human CD25 Positive Selection Kit Protocol (see Table 1), the negative fraction can be further depleted of CD25+ cells to obtain CD4+CD25- cells. Depending on the donor, purity of CD4+ T cells in the CD25-depleted fraction may vary.

See page 1 for Recommended Medium. Refer to Table 2 for detailed instructions regarding the depletion procedure.

**Table 2. EasySep™ Human CD25 Depletion Kit Protocol**

|      |   | EASYSEP™ MAGNETS  |   |
|------|---|---|---|
| STEP | INSTRUCTIONS  |  <b>EasySep™</b><br>(Catalog #18000) | <b>“The Big Easy”</b><br>(Catalog #18001)  |
| 1    | Centrifuge sample and resuspend in the indicated volume of recommended medium.  | 0.5 mL  | 0.5 mL  |
|      | Add sample to required tube.  | 5 mL (12 x 75 mm) polystyrene round-bottom tube<br>(e.g. Corning Catalog #352058)                                     | 14 mL (17 x 100 mm) polystyrene round-bottom tube<br>(e.g. Corning Catalog #352057)   |
| 2    | Add Selection Cocktail to sample.   | 50 µL   | 50 µL   |
|      | Mix and incubate.   | RT for 15 minutes   | RT for 15 minutes   |
| 3    | Mix Magnetic Particles.   | Pipette up and down more than 5 times   | Pipette up and down more than 5 times   |
| 4    | Add Magnetic Particles to sample.   | 50 µL   | 50 µL   |
|      | Mix and incubate.   | RT for 10 minutes   | RT for 10 minutes   |
| 5    | Add recommended medium to top up the sample to the indicated volume. Mix by gently pipetting up and down 2 - 3 times.               | Top up to 2.5 mL  | Top up to 2.5 mL  |
| 6    | Place the tube (without lid) into the magnet and incubate.  | RT for 10 minutes   | RT for 10 minutes   |
| 7    | Pick up the magnet, and in one continuous motion invert the magnet and tube,* pouring the enriched cell suspension into a new tube. | Use a new 5 mL tube   | Use a new 14 mL tube  |
| 8    | Remove the tube from the magnet and place the new tube (without lid) into the magnet and incubate for a second separation.          | RT for 10 minutes   | RT for 10 minutes   |
| 9    | Pick up the magnet, and in one continuous motion invert the magnet and tube,* pouring the enriched cell suspension into a new tube. | Isolated cells are ready for use  | Isolated cells are ready for use  |


RT - room temperature (15 - 25°C)

\* Leave the magnet and tube inverted for 2 - 3 seconds, then return upright. Do not shake or blot off any drops that may remain hanging from the mouth of the tube.

## Directions for Use – Fully Automated RoboSep™ Protocol

See page 1 for Recommended Medium. Refer to Table 3 for detailed instructions regarding the RoboSep™ procedure.

**Table 3. RoboSep™ Human CD25 Positive Selection Kit Protocol**

| STEP | INSTRUCTIONS  | RoboSep™<br>(Catalog #20000 and #21000)   |  |
|------|---|---|---|
| 1    | Prepare sample at the indicated volume.   | 0.5 mL  |   |
|      | Add sample to required tube.  | 14 mL (17 x 100 mm) polystyrene round-bottom tube<br>(e.g. Corning Catalog #352057) |   |
| 2    | Select protocol.  | • Human CD25high Positive Selection 18231-High Purity                               |   |
| 3    | Mix Magnetic Particles.   | Pipette up and down more than 5 times   |   |
| 4    | Load the carousel.  | Follow on-screen prompts  |   |
|      | Start the protocol.   | Press the green “Run” button  |   |
| 5    | Unload the carousel when the run is complete. Remove the tube containing the isolated cells and resuspend in desired medium. Be sure to collect cells from the sides of the tube. | Isolated cells are ready for use  |   |

## Notes and Tips

### ASSESSING PURITY

EasySep™ Human CD25 Positive Selection Cocktail uses an anti-CD25 antibody clone which recognizes epitope B of the CD25 antigen and may block some anti-CD25 antibody clones used to assess purity by flow cytometry. For purity assessment by flow cytometry use the following fluorochrome-conjugated antibody clones:

- Anti-Human CD25 Antibody, Clone 2A3, PE (Catalog #60153), or clone BC96 which recognize epitope A of the CD25 antigen

The following method can also be used:

- Use a fluorochrome-conjugated secondary antibody, such as Goat Anti-Mouse IgG (H+L) Antibody, Polyclonal (Catalog #60138).

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