



## EasySep™ Mouse Monocyte Enrichment Kit

Negative Selection

Catalog #19761

For processing  $1 \times 10^9$  cells



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## Description

Isolate untouched and highly purified monocytes from mouse bone marrow or peripheral blood samples by immunomagnetic negative selection. When using single-cell suspensions from other tissue types, this kit may require optimization.

- Fast, easy-to-use and column-free
- Up to 98% purity (blood), 93% purity (bone marrow)
- Isolated cells are untouched

This kit targets non-monocytes for removal with biotinylated antibodies recognizing specific cells surface markers. Unwanted cells are labeled with biotinylated antibodies and magnetic particles, and separated without columns using an EasySep™ magnet. Desired cells are simply poured off into a new tube. Isolated cells are immediately available for downstream applications such as flow cytometry, culture, or DNA/RNA extraction.

## Component Descriptions

COMPONENT NAME	COMPONENT #	QUANTITY	STORAGE	SHELF LIFE	FORMAT
EasySep™ Mouse Monocyte Enrichment Cocktail	19761C.1	1 x 1 mL	Store at 2 - 8°C. Do not freeze.	Stable until expiry date (EXP) on label.	A combination of monoclonal antibodies in PBS and 0.1% BSA.
EasySep™ Biotin Selection Cocktail	19153	1 x 1 mL	Store at 2 - 8°C. Do not freeze.	Stable until expiry date (EXP) on label.	A combination of monoclonal antibodies in PBS.
EasySep™ D Magnetic Particles	19250	2 x 1 mL	Store at 2 - 8°C. Do not freeze.	Stable until expiry date (EXP) on label.	A suspension of magnetic particles in TBS.
Normal Rat Serum	13551	1 x 2 mL	Store at -20°C	Stable until expiry date (EXP) on label.	Mycoplasma-free normal rat serum.

BSA - bovine serum albumin; PBS - phosphate-buffered saline; TBS - TRIS-buffered saline

Components may be shipped at room temperature (15 - 25°C) but should be stored as indicated above.

## Additional Reagent Stability Information

REAGENT NAME	STORAGE	SHELF LIFE
Normal Rat Serum (in-use)	Store at 2 - 8°C.	Stable for at least 2 months. Do not exceed expiry date (EXP) on label.

## Sample Preparation

### BONE MARROW

Flush bone marrow cells from femur and tibia into recommended medium using a syringe equipped with a 23 gauge needle. Disperse clumps by gently passing the cell suspension through the syringe several times. Alternatively, crush bones using a mortar and pestle. Remove remaining clumps and debris by passing cell suspension through a 70 µm mesh nylon strainer. Centrifuge at 300 x g for 10 minutes and resuspend cells at  $1 \times 10^8$  cells/mL in recommended medium.

### PERIPHERAL BLOOD

Blood should be lysed prior to use. Mix 1 part blood with 9 parts Ammonium Chloride Solution (Catalog #07800) and incubate on ice for 15 minutes. Centrifuge at 300 x g for 6 minutes. Discard supernatant and wash cell pellet once with recommended medium. Discard supernatant and resuspend cell pellet at  $1 \times 10^8$  cells/mL in recommended medium.

If there are less than  $5 \times 10^7$  cells/mL, resuspend in 500 µL of recommended medium.



## Recommended Medium

EasySep™ Buffer (Catalog #20144), RoboSep™ Buffer (Catalog #20104), or PBS containing 2% fetal bovine serum (FBS) and 1 mM EDTA. HBSS, Modified (Without Ca++ and Mg++; Catalog #37250) can be used in place of PBS. Medium should be free of Ca++ and Mg++.

## Directions for Use – Manual EasySep™ Protocols

See page 1 for Sample Preparation and Recommended Medium. Refer to Table 1 for detailed instructions regarding the EasySep™ procedure for each magnet.

**Table 1. EasySep™ Mouse Monocyte Enrichment Kit Protocol**

		EASYSEP™ MAGNETS	
STEP	INSTRUCTIONS	 <b>EasySep™</b> (Catalog #18000)	 <b>“The Big Easy”</b> (Catalog #18001)
1	Prepare sample at the indicated cell concentration within the volume range.	1 x 10 <sup>8</sup> cells/mL 0.5 - 2 mL	1 x 10 <sup>8</sup> cells/mL 0.5 - 6 mL
2	Add Rat Serum to sample.	50 µL/mL of sample	50 µL/mL of sample
3	Add sample to required tube.	5 mL (12 x 75 mm) polystyrene round-bottom tube (e.g. Corning Catalog #352058)	14 mL (17 x 100 mm) polystyrene round-bottom tube (e.g. Corning Catalog #352057)
4	Add Enrichment Cocktail to sample.	50 µL/mL of sample	50 µL/mL of sample
	Mix and incubate.	2 - 8°C for 15 minutes	2 - 8°C for 15 minutes
5	Wash the cells by topping up the sample tube with recommended medium and centrifuging.	300 x g for 10 minutes	300 x g for 10 minutes
	Discard the supernatant and resuspend cells in the original volume with recommended medium.	0.5 - 2 mL	0.5 - 6 mL
6	Add Biotin Selection Cocktail to sample.	60 µL/mL of sample	60 µL/mL of sample
	Mix and incubate.	2 - 8°C for 15 minutes	2 - 8°C for 15 minutes
7	Vortex Magnetic Particles. NOTE: Particles should appear evenly dispersed.	30 seconds	30 seconds
8	Add Magnetic Particles to sample.	150 µL/mL of sample	150 µL/mL of sample
	Mix and incubate.	2 - 8°C for 10 minutes	2 - 8°C for 10 minutes
9	Add recommended medium to top up the sample to the indicated volume. Mix by gently pipetting up and down 2 - 3 times.	Top up to 2.5 mL	<ul style="list-style-type: none"> <li>• Top up to 2.5 mL for samples ≤ 2 mL</li> <li>• Top up to 10 mL for samples &gt; 2 mL</li> </ul>
	Place the tube (without lid) into the magnet and incubate.	RT for 5 minutes	RT for 5 minutes
10	Pick up the magnet, and in one continuous motion invert the magnet and tube,* pouring the enriched cell suspension into a new tube.	Isolated cells are ready for use	Isolated cells are ready for use


RT - room temperature (15 - 25°C)

\* Leave the magnet and tube inverted for 2 - 3 seconds, then return upright. Do not shake or blot off any drops that may remain hanging from the mouth of the tube.

## Directions for Use – Fully Automated RoboSep™ Protocol

See page 1 for Sample Preparation and Recommended Medium. Refer to Table 2 for detailed instructions regarding the RoboSep™ procedure.

**Table 2. RoboSep™ EasySep™ Mouse Monocyte Enrichment Kit Protocol**

STEP	INSTRUCTIONS	RoboSep™ (Catalog #20000 and #21000)	
1	Prepare sample at the indicated cell concentration within the volume range.	1 x 10 <sup>8</sup> cells/mL 0.5 - 6 mL	
2	Add Rat Serum to sample.	50 µL/mL of sample	
3	Add sample to required tube.	14 mL (17 x 100 mm) polystyrene round-bottom tube (e.g. Corning Catalog #352057)	
4	Add Enrichment Cocktail to sample.	50 µL/mL of sample	
	Incubate.	2 - 8°C for 15 minutes	
5	Wash the cells by topping up the sample tube with recommended medium and centrifuging.	300 x g for 10 minutes	
	Discard the supernatant and resuspend cells in the original volume with recommended medium.	0.5 - 6 mL	
6	Select protocol.	Mouse Monocyte Negative Selection 19761	
7	Vortex Magnetic Particles. NOTE: Particles should appear evenly dispersed.	30 seconds	
8	Load the carousel.	Follow on-screen prompts	
	Start the protocol.	Press the green "Run" button	
9	Unload the carousel when the run is complete.	Isolated cells are ready for use	

## Notes and Tips

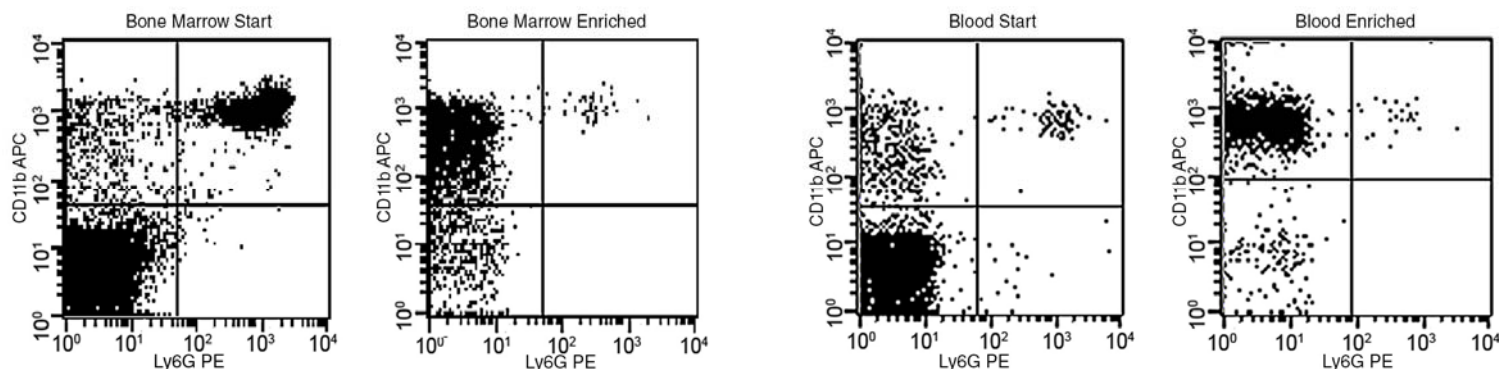
### ASSESSING PURITY

To date, an exclusive marker for mouse bone marrow and peripheral blood monocytes has not been identified. However, monocytes are known to express CD11b, F4/80, CD115 (M-CSFR), Gr-1, and Ly-6C.

For purity assessment of monocytes (CD11b+Ly-6G-) by flow cytometry use the following fluorochrome-conjugated antibody clones:

- Anti-Mouse CD11b Antibody, Clone M1/70 (Catalog #60001), and
- Anti-Mouse Ly-6G Antibody, Clone 1A8 (Catalog #60031)

## Data



Starting with mouse bone marrow or peripheral blood, the monocyte content (CD11b+Ly-6G-) of the enriched fraction typically ranges from 92 - 98% (blood) and 80 - 93% (bone marrow). In the above examples, the purities of the start and final enriched fractions are 8.8% and 88.8% (bone marrow), and 9.7% and 95.1% (blood), respectively.

NOTE: RBCs were removed by lysis prior to flow cytometry.

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